

## DATA SHEET

**Hot Start Taq Pol Master Mix (2X)**

Master mix of Hot Start Taq Polymerase

Cat. Nº.	Amount
<input type="checkbox"/> POL-116XS	50 reactions
<input type="checkbox"/> POL-116S	100 reactions
<input type="checkbox"/> POL-116M	200 reactions
<input type="checkbox"/> POL-116L	500 reactions
<input type="checkbox"/> POL-116XL	1.000 reactions

**Shipping:**

Shipped on blue ice

**Storage Conditions:**

Store at -20 °C

**Additional Storage Conditions:**

Avoid freeze/thaw cycles. Hot Start Taq Pol Master Mix (2X) is also stable for three months at 4°C, so for frequent use, an aliquot may be kept at 4°C.

**Shelf Life:**

12 months

**For *in vitro* use only!****Description:**

Hot Start Taq Pol Master Mix contains Hot Start Taq Polymerase in an optimized PCR buffer with Mg<sup>2+</sup> and dNTPs. It contains all reagents required for PCR (except template and primer) in a premixed 2x concentrated ready-to-use solution. The Master Mix is recommended for use in routine PCR reactions. It is optimized for high specificity and guarantees minimal by-product formation. Antibody-based hot start technology avoids nonspecific amplification and enables room temperature reaction setup. The engineered Taq DNA Pol hot start enzyme allows amplification of fragments up to 5 kbp.

**Kit contents:****2x Hot Start Taq Pol Master Mix (purple cap)**

Master mix of thermostable Hot Start DNA polymerase, dATP, dCTP, dGTP, dTTP, KCl, MgCl<sub>2</sub> and stabilizers.

**PCR Reaction Setup**

Use the quantities below to prepare a single 50 µl PCR reaction. Thaw, mix, and briefly centrifuge each component before use. Add the following components to a microcentrifuge tube:

**1. Prepare PCR master mix**

Note: Consider the volumes for all components listed next steps to determine the correct amount of water required to reach your final reaction volume.

Components	50 µL rxn	[ final ]
Water, grade PCR	To 50 µl	
2 X Hot Start Taq Pol Master Mix	25 µl	1X

Mix and briefly centrifuge the components.

**2. Add template DNA and primers**

Components	50 µL rxn	[ final ]
Foward primer (10 µM)	0,5 - 2,5 µl	0,1 – 0,5 µM
Reverse primer (10 µM)	0,5 - 2,5 µl	0,1 – 0,5 µM
DNA template		10 pg – 1 µg**

\*\*genomic DNA: 1 ng-1µg; plasmidial ou viral DNA: 1 pg-1 ng

Cap each tube, mix, and briefly centrifuge the content.

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**3. Incubate reactions in a thermal cycler.**

Recommended cycling conditions:

Step	Temp.	Time
Initial denaturation	95 °C	1 min
30 cycles	Denaturation	95 °C 15 - 30 sec
	Annealing <sup>1</sup>	45-68 °C 15 - 30 sec
	Elongation <sup>2</sup>	72 °C 30 sec - 4 min
Final extension	72 °C	2 min
Hold	4 - 8 °C	

1)The annealing temperature depends on the melting temperature of the primers used.

2)The elongation time depends on the length of the fragments to be amplified. A time of 1 min/kb is recommended.

For optimal specificity and amplification an individual optimization of the recommended parameters may be necessary for each new template DNA and/or primer pair.